TML/MSH Microbiology Department	Policy # MI/VIR/16/20/v02	Page 1 of 2
Policy & Procedure Manual	-	
Section: Virology Manual	Subject Title: Appendix XX	
	Cytospin Preparation	
Issued by: LABORATORY MANAGER	Original Date: June 25, 2002	
Approved by: Laboratory Director	Revision Date: October 10, 2003	

Appendix XX

CYTOSPIN PREPARATION

I. Introduction

A cytospin preparation is a concentration of cells taken directly from specimens or from scraped cell cultures.

II. Reagents and Materials

Virus-specific or pooled antibody Phosphate buffered saline (PBS) Cold acetone (4°C) Distilled water Mounting fluid Non-immune antibody vortex sterile pipettes cytospin and accessories humidified chamber coplin jars fluorescence microscope

III. Procedure

1. Shell Vial

- i. Remove all except 1 ml maintenance media from shell vial using a sterile pipette.
- ii. Scrape cells from top of coverslip using a sterile pipette. Break up cell clumps by pipetting the cells up and down several times.
- iii. Pipette 200 ul (4 drops) of scraped cells into funnel for each well.
- iv. Cytospin at 2000 rpm (700g) for 5 minutes.
- v. Remove slide and air dry.
- vi. Fix in cold acetone for 10 minuets in a coplin jar. Remove slide and air dry.

 $PROCEDURE\ MANUAL \\ TORONTO\ MEDICAL\ LABORATORIES\ /\ MOUNT\ SINAI\ HOSPITAL\ MICROBIOLOGY\ DEPARTMENT$

TML/MSH Microbiology Department	Policy # MI/VIR/16/20/v02	Page 2 of 2
Policy & Procedure Manual		
Virology Manual		

vii. Proceed to staining. Refer to Appendix IV for Indirect fluorescent antibody staining techniques or Appendix V for Direct fluorescent antibody staining techniques.

or

Refer to Appendices IV and V for immunofluorescent staining techniques for shell vials.

2. Tube culture

- i. Remove all except 1 ml maintenance media from the culture tube using a sterile pipette.
- ii. Scrape cells from side of tube using a sterile pipette. Break up cell clumps by pipetting the cells up and down several times.
- iii. Pipette 200 ul (4 drops) of scraped cells into funnel for each well.
- iv. Cytospin at 2000 rpm (700 x g) for 5 minutes.
- v. Remove slide and air dry.
- vi. Fix in cold acetone for 10 minutes in a coplin jar. Remove slide and air dry.
- vii. Proceed to staining. Refer to Appendix IV for Indirect fluorescent antibody stains or Appendix V for Direct fluorescent antibody stains.

or

Refer to Appendices IV and V for immunofluorescent staining techniques for shell vials.

3. Direct from specimen

IV. Reference

Thermo Shandon, cytospin. Manufacturer's manual. Refer to Appendix VI for procedure.