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Section: Parasitology Manual	Subject Title: Quality Assurance Procedures for Parasitology	
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QUALITY ASSURANCE PROCEDURES FOR THE PARASITOLOGY LABORATORY

1. Equipment

Microscopes:

- The microscopes located in Parasitology are used exclusively by those technologists and are under a service contract under which all microscopes are serviced yearly.
- It is the responsibility of the technologist using the microscope to check the Kohler illumination before starting the day's work and to record and initial this procedure in the Kohler illumination binder.
- It is essential that objectives not be moved between microscopes since it may affect the calibration of the microscopes.
- The condenser's numerical aperture should be equal or greater than the highest objective numerical aperture.
- Ensure that spare light bulbs are available for all microscopes
- use a camel hair brush to clean eyepieces and all glass surfaces
use disinfectant (70% isopropyl alcohol) to clean the stage
- keep microscopes covered when not in use.
- report any frayed or broken electrical wires.

Centrifuges:

- before each run visually inspect the device for loose or damaged parts
- inspect any seals and O-rings on buckets and lids
- inspect the interior for signs of leaking on a previous run and clean with disinfectant as required
- disinfect the centrifuge monthly
- the speed can be checked with a stroboscope or tachometer quarterly
- check the brushes every six months
- record any problems encountered

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Fume Hood:

- at least yearly check to make sure the hood draws air in at a velocity of 100ft/min with the sash fully open
- lubricate the sash guides
- ensure that the minimum amount of material is being stored in the hood
- ensure that stored material does not impede air circulation

Biological Safety Cabinet:

- The BSC is under contract to be cleaned and recertified once a year (see sticker on the cabinet).
- after each use disinfect the work area (no UV light is provided, use bleach solution)
- if the cabinet is moved it must be decontaminated and recertified.
- do not open any section of the cabinet
- check the air pressure gauge on the front of the cabinet. Do not use if the gauge indicates a pressure outside of the operating limits (see gauge) and record reading daily on sheet on side of cabinet.

2. Control slides

- Control slides are used in the various staining procedures performed in the lab.
- Control slides are to be used with each rack of slides to be stained for the Sequential Stain (routine lab stain), Hematoxylin Stain, Modified Kinyoun Stain and Modified Trichrome Stain (Chromotrope Stain).
- The control slides are read by the technologist and the results are recorded and initialled on record sheets maintained in the Laboratory Manual.
- Specimens received for testing such as QMP-LS are handled in a routine manner and are read by the assigned technologist.
- Staining control slides are used whenever QMP-LS specimens are stained.
- The following quality control organisms are available from the ATCC :
 - ATCC 30010 *Acanthamoeba castellanii*
 - ATCC 30133 *Naegleria gruberi*
 - ATCC 30925 *Entamoeba histolytica*
 - ATCC 30001 *Trichomonas vaginalis*
 - ATCC 30883 *Leishmania mexicana*
 - ATCC 30160 *Trypanosoma cruzi*
 - *Escherischei coli* can be obtained from the Microbiology Dept.

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3. Participation in Quality Assurance Programs:

- We participate in QMP-LS, which provides stool samples for preparation, staining and reading.
- All positive malaria bloods are kept and are frozen for PCR testing. Malaria speciation can then be confirmed by this accepted gold standard.
- Monthly meetings are held to discuss laboratory related issues and to determine if an evaluation of a new method is warranted.
- Participation in both the CAP Parasitology and the CAP Blood Parasite surveys.